# **Xylitol Dehydrogenase**

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# **Summary**

Xylitol dehydrogenase catalyzes the oxidation of xylitol to xylulose using NAD<sup>+</sup> as a co-substrate. This protocol describes a direct enzyme assay for determining xylitol dehydrogenase activity. This assay is only suitable for those cases in which a competing enyzme reaction, the oxidation of xylitol to xylose, uses NADP<sup>+</sup> as a co-substrate.

# Solutions Required

- 1. 500 mM tris-HCl buffer pH = 8.6. adjust pH to 8.6 with 20% KOH.
- 2. 10 mM potassium phosphate buffer pH = 7.0 prepared by mixing 8 mL of 10 mM KH<sub>2</sub>PO<sub>4</sub> and 20 mL of 10 mM K<sub>2</sub>HPO<sub>4</sub> or prepared by dissolving 0.0194 g KH<sub>2</sub>PO<sub>4</sub> and 0.0815 g K<sub>2</sub>HPO<sub>4</sub>·3H<sub>2</sub>O in 50 mL water.
- 3. 100 mM 2-mercaptoethanol can be prepared in stock solution and stored in refrigerator.
- 4. 4.0 mM NAD<sup>+</sup> must be prepared fresh
- 5. 1.5 M xylitol can be prepared in stock solution and stored in refrigerator.

# Preparation of Cell Extract

Follow general protocol **Preparation of Cell Extract**.

- 1. After first pelletization of cells, resuspend at 4°C in potassium phosphate buffer.
- 2. After second pelletization of cells, resuspend at 4°C in potassium phosphate buffer.

# **Spectrophotometer**

Turn on the ultraviolet bulb on the spectrophotometer (Beckman DU50) and wait 10 minutes for warm-up. Select the kinetics-time window on the instrument. Load the method "A:/nadh30" or "A:/nadh37". These methods each have a run-time of 60 s, a temperature of 30°C or 37°C (respectively), a wavelength of 340 nm and use 2 autosamplers.

### Procedure

1. For each assay, prepare the two cocktails shown in the following table into two separate UV-translucent cuvettes, and keep them on ice.

	Volume (µL) added to:	
Solution	Control	Experimental
DI H <sub>2</sub> O	300	200
500 mM tris-HCl	400	400
2-mercaptoethanol	100	100
$\mathrm{NAD}^{^{+}}$	100	100
xylitol	0	100

- 2. Directly from the ice when ready to commence the assay, place the two quartz cuvettes (each containing 950 µL) into the spectrophotometer holder (position #1 for control, position #2 for experimental). Use cuvette lid caps to mix 3 or 4 times then insert in instrument.
- 3. Wait 10 minutes to allow the temperature of the solutions in the cuvettes to equilibrate.
- 4. "Blank" and then depress "Read Samples" on the monitor.
- 5. Simultaneously add  $100 \,\mu\text{L}^{\dagger}$  of the cell extract to the cuvettes.
- 6. To mix solutions, immediately and simultaneously aspirate and dispense the contents of the cuvettes with a pipettor. Mix the solutions in this way ten times. (Count!)
- 7. Promptly depress "start" on the monitor.
- 8. Record the rates for the two (control and experimental) cuvettes.

† Dilution of the cell extract may be adjusted so that change in absorbance is between about 0.05 and 0.7 AU in one minute. This dilution should be accomplished externally in a microcentrifuge tube (for example, by adding 50  $\mu$ L of cell extract to 950  $\mu$ L DI water to achieve a dilution of 20). The volume of 100  $\mu$ L should always be used in the enzyme assay mixture.

# Calculation of Activity

One unit (U) of xylitol dehydrogenase activity is defined as the amount of enzyme required to oxidize 1.0 µmole of xylitol in one minute.

1. 
$$dA/dt (min^{-1}) = [Rate]_{experimental} - [Rate]_{control} = dA/dt$$

2. Activity = 
$$\frac{1000 \times TV \times D \times dA/dt}{\varepsilon \times V \times CF}$$

Activity: Volumetric Activity (U/L)

TV: Total volume in cuvette (1000 µL)

D: Dilution of the cell extract. (For example, if 50 µL of cell extract were add to 950 µL

DI water prior to using a volume of cell extract in the assay, then D=20)

V: Volume of cell extract used (50 µL)

E: Molar extinction coefficient for NADH (6.22 L/mmol for a path length of 1.0 cm)

CF: Concentration Factor of cell extract (For example, if a 100 mL sample is

concentrated to a 2 mL volume for the French Press, then CF=50)

3. Specific Activity = 
$$\frac{Activity}{Protein Concentration}$$
 1

Activity: Volumetric Activity, as calculated in #2 above (U/L)

Protein Concentration: Protein concentration, as calculated in protocol Total Protein

**Concentration** (mg/L)

Specific Activity: (U/mg protein)

# <u>Reference</u>

T. Ikeuchi, R. Kiritani, M. Azuma, H. Ooshima (2000) Effect of D-glucose on induction of xylose reductase and xylitol dehydrogenase in *Candida tropicalis* in the presenceof NaCl J. Basic Microbiol. 40(3), 167-175.